# **T400-Series Surgical Protocol**

## Mouse Femoral Artery: Chronic Blood Flow Measurement

#### **APPLICATION BASICS**

Site: Femoral artery

Species: Mouse
Body Weight: 17 - 30 grams
Duration: Chronic
Vessel Diameter: 0.20 - 0.40 mm

Probe

Size: 0.5 mm

Reflector: J Connector: 4-pin Cable Length: 10 cm

Catalog #: MC-0.5PSS-JN-WC10-CA4S-GC TS420 Perivascular Module

### **Anatomical Landmarks**

The femoral artery lies distal to the epigastric artery. The segment of the femoral artery proximal to the epigastric artery is typically isolated for acute or chronic flow measurements (Fig.1). The vessel at this point measures ~250-300  $\mu m$  in 25 g CD-1 mice. Small branches from the femoral are typically found at both locations where the 7-0 silk passes under the femoral artery.

Isolation of the artery from the vein in this position is very challenging. There is less fascia and connective tissue around the sheath of the artery and vein to manipulate the vessel free. The nerve runs immediately adjacent and is very easy to tear. Once isolated, avoid pulling up on the artery which would cause vascular spasm or putting pressure on the vein to cause occlusion.

The protocol presents an overview of steps to achieve successful femoral arterial blood flow measurement in the mouse adapted from the Considerations for Chronic Flow Measurement on the Mouse Femoral Artery PowerPoint presentation. Courtesy of Michael F. Callahan, Dept. of Orthopaedic Surgery, Wake Forest University School of Medicine, Winston-Salem, NC. For the complete photo essay PowerPoint presentation see:

www.transonic.com

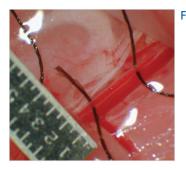


Fig. 1: The femoral artery proximal to the epigastric artery, isolated from the femoral vein, measures ~400 µm diameter with a micrometer. Flow measurements can be made at this location fairly easily.

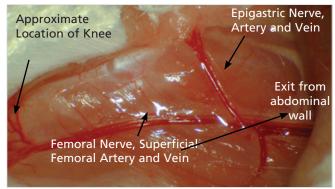


Fig. 2: Figure shows femoral nerve, superficial femoral artery and femoral vein exiting from abdominal wall and running down the leg toward the knee. Note position of epigastic nerve, artery and vein.



Fig. 3: The superficial femoral artery, distal to the epigastric artery, measures ~200 µm diameter and is very challenging to isolate without damaging the nerve or the vein.



Fig. 4: A 0.5PSS Flowprobe prepared with dacron mesh attached to the underside of the Probe with silicone adhesive.



# Mouse Femoral Artery: Chronic Blood Flow Measurement Cont.

### **Before Surgery**

Prepare a 0.5PSS Flowprobe with a dacron mesh attached to the underside of the Probe with silicone adhesive to help stabilize the Probe position in the implant (Fig. 4).

#### Chronic Measurement Protocol cont.

#### 1. Femoral Artery Isolation

(Acute or Chronic Flow Measurements. Refer to Figs. 1-3)

The segment of the femoral artery proximal to the epigastric artery is typically isolated for acute or chronic flow measurements. The vessel at this point measures ~250-300  $\mu m$  in 25 g CD-1 mice. Small branches from the femoral are typically found at both locations where the 7-0 silk passes under the femoral artery.

#### 2. Thigh Incision

Prepare the medial thigh and a 2 cm area on the back for surgery. A 1.5 cm incision is made into the medial aspect of the thigh near the body wall (Fig. 5). Care should be taken to avoid the epigastric artery which exits the femoral about 7-8 mm from the body wall and travels rostrally in a fat pad attached to the skin.

#### 3. Positioning Probe Connector

Make an incision in the back to place the Flowprobe connector and then tunnel subcutaneously from the back to the posterior aspect of the thigh (Fig. 6). With a pair of forceps pull the Probe connector from the back of the thigh to the exit on the back. The length of the cable was short in this prototype Probe and prevented placement in the preferred midscapulae position. Therefore, the connector was placed on the small of the back.

#### 4. Isolation of the Femoral Artery

Open the sheath covering the femoral artery, vein and nerve with a pair of 45° microblunted Dumont forceps parallel to the vessel. Typically, one small branch is located between the abdominal wall and the epigastric artery. When the fascia is removed, a 3-4 mm section of the artery between the small branch and the epigastric will appear (Fig. 7).

Clear this section to place

the Flowprobe. Grasp the fat/fascia located rostral to

the vessels and pull it rostrally. The nerve can then be

isolated from the vessels with a 45° Dumont without

touching or damaging it (Fig. 8). Pulling on the fascia caudal to the vein allows it to be separated from the artery. A small branch of the femoral can usually be

proximal to the epigastric artery (Fig. 9).

seen exiting caudally on the femoral artery immediately

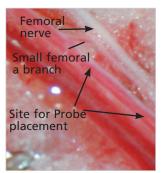


Fig. 8: Nerve isolated from the femoral vessels.

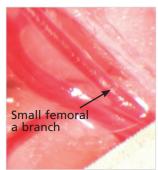


Fig. 9: A small branch of the femoral exiting caudally on the femoral a. proximal to the epigastric a.



Fig. 5: Incision in medial aspect of thigh.



Fig. 6: An incision is made in the back to place the Flowprobe connector.

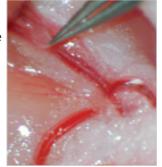


Fig. 7: Femoral artery isolated.



## Mouse Femoral Artery: Chronic Blood Flow Measurement Cont.

#### Chronic Measurement Protocol cont.

#### 5. Placement and Securing of the Flowprobe

Gently place the Probe in position over the vessels and pull the dacron mesh under the artery (Fig. 10). The Probe cable and or mesh can be anchored to the muscle with 7-0 silk suture prior to passing the mesh under the vessel but these anchor points should be placed before positioning the Probe.

After passing the mesh, gently lift the femoral artery and place it into the lumen of the Probe. Place a section of dacron mesh over the Probe and secure the two pieces of mesh with silk (Fig. 11). Place acoustic coupling gel or lidocaine gel into the lumen of the Probe to displace air bubbles. The gel acts as a barrier to prevent the Kwik-Sil from entering the lumen of the Probe. Check the signal of the Flowprobe on the Flowmeter.

Inject 2-part Kwik-Sil or Kwik-Cast (shown) around the Flowprobe and cable (Fig. 12). The vessel can be visualized under the mesh, exiting the lumen of the Probe. Do not inject the casting material into the lumen of the Probe. This will block the ultrasound signal, and prevent flow measurement. Allow the cast to become hardened before closing wounds. Prepare skin button as previously indicated.

#### 6. Experimental Exercise Setup

The mouse is connected to a Flowmeter via an electronic swivel connected to the skin button (Fig. 13). Femoral arterial blood flow is recorded by the Flowmeter as the mouse, implanted with a Nanoprobe on the femoral artery, runs on the treadmill.

### 7. Flowprobe Recovery After 5 Weeks Implantation

Connective tissue and blood vessels overlie the Kwik-Cast but have not infiltrated it (Fig. 16). Tissue has infiltrated the mesh but this can be cut away (Fig. 17) to visualize the intact blood vessel exiting the Probe lumen (right). At this point one can check for "zero" flow by occluding the vessel.

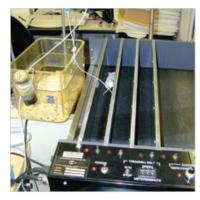


Fig. 13: Three station treadmill for rodents by Columbus Instruments.



Fig. 14: Mouse with femoral Nanoprobe implant running on treadmill.



Fig. 10: Dacron mesh pulled under the artery.



Fig. 11: Upper and lower pieces of Dacron mesh secured with two pieces of silk.



Fig. 12: Kwik-Sil or Kwik-Cast injected around the Probe and cable.



# Mouse Femoral Artery: Chronic Blood Flow Measurement Cont.

#### Chronic Measurement Protocol cont.

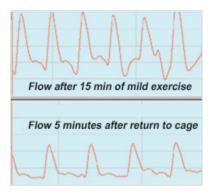


Fig. 15: Effect of brief exercise on femoral artery blood flow.

Mean Flows: Top: 1.9 ml/min during treadmill exercise;

Bottom: 0.8 ml/min returned to home cage.



Fig. 16: Kwik-Cast with connective tissue and blood vessels.

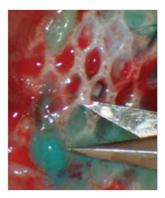


Fig. 17: Cutting away tissues from the Dacron mesh.



Fig. 18: Probe removed.

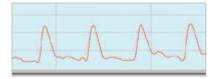


Fig. 19: Typical mouse chronic femoral artery flowtrace.

#### **ACKNOWLEDGEMENTS**

Transonic Systems Inc.® gratefully acknowledges the assistance of Michael F. Callahan, PhD, and Thomas L. Smith, PhD, Department of Orthopaedic Surgery, Wake Forest University School of Medicine, Winston-Salem, NC, in the development of this protocol.

Adapted from a PowerPoint presentation produced by: Margo Sosa, Senior Product Manager, Transonic Systems, Inc., 34 Dutch Mill Road, Ithaca, NY 14850 www.transonic.com

#### **REFERENCES**

Cabou C Campistron G, Marsollier N, Leloup C, Cruciani-Guglielmacci C, Pénicaud L, Drucker DJ, "Brain glucagon-like peptide-1 regulates arterial blood flow, heart rate, and insulin sensitivity," Diabetes 2008; 57(10): 2577-87.

Cabou C, Cani PD, Campistron G, Knauf C, Mathieu C, Sartori C, Amar J, Scherrer U, Burcelin R, "Central insulin regulates heart rate and arterial blood flow: an endothelial nitric oxide synthase-dependent mechanism altered during diabetes," Diabetes 2007; 56(12): 287.



Transonic Systems Inc. is a global manufacturer of innovative biomedical measurement equipment. Founded in 1983, Transonic sells "gold standard" transit-time ultrasound flowmeters and monitors for surgical, hemodialysis, pediatric critical care, perfusion, interventional radiology and research applications. In addition, Transonic provides pressure and pressure volume systems, laser Doppler flowmeters and telemetry systems.

#### **AMERICAS**

Transonic Systems Inc. 34 Dutch Mill Rd Ithaca, NY 14850 U.S.A.

Tel: +1 607-257-5300 Fax: +1 607-257-7256 support@transonic.com

#### **EUROPE**

Transonic Europe B.V.
Business Park Stein 205
6181 MB Elsloo
The Netherlands
Tel: +31 43-407-7200
Fax: +31 43-407-7201
europe@transonic.com

#### ASIA/PACIFIC

Transonic Asia Inc. 6F-3 No 5 Hangsiang Rd Dayuan, Taoyuan County 33747 Taiwan, R.O.C. Tel: +886 3399-5806 Fax: +886 3399-5805 support@transonicasia.com

#### **JAPAN**

Transonic Japan Inc. KS Bldg 201, 735-4 Kita-Akitsu Tokorozawa Saitama 359-0038 Japan Tel: +81 04-2946-8541 Fax: +81 04-2946-8542 info@transonic.jp